Biota Vol. XI (3): 180-184, Oktober 2006 ISSN 0853-8670

Genetic Characterstics of the Kintamani Dogs Using Microsatellite Markers

Karakterisasi Genetik Anjing Kintamani Menggunakan Petanda Mikrosatelit

I Ketut Puja

Faculty of Veterinary Medicine, Udayana University, Denpasar Bali E-mail: asubali@hotmail.com

Abstrak

Karakterisasi molekuler anjing Kintamani berperan penting dalam penentuan status trah dan mencegah penurunan kemurnian akibat perkawinan silang dengan trah anjing lain. Anjing Kintamani sangat popular di Bali. Anjing Kintamani dipercaya berasal dari Kintamani, Bangli, tetapi asal usulnya sampai kini belum jelas. Mikrosatelit merupakan runutan nukleotida terulang yang didistribusikan secara acak dalam gen vertebrata. Lokus mikrosatelit ini telah diketahui sangat polimorfik, karena keragaman jumlah nukleotida yang terulang. Mikrosatelit merupakan petanda allel utama yang dapat digunakan untuk pemetaan gen, genetika populasi dan identifikasi individu serta alat untuk mengungkap karakter genetik hewan. Pada penelitian ini diungkap karakteristik genetik anjing Kintamani menggunakan mikrosatelit. Jumlah dan ukuran allel pada 425 anjing Kintamani dianalisis menggunakan 116 macam primer mikrosatelit. DNA diisolasi dari sel hasil swab pipi. Amplifikasi 116 lokus mikrosatelit menggunakan PCR dalam 12 multiplek. Produk PCR dipisahkan dengan gel bis-akrilamid 6% dalam automated DNA sequencer. Flurosesnsi yang dihasilkan dideteksi dengan program Genescan 3.1, dan program Strand versi 2.2.39 digunakan untuk menghitung jumlah allel. Hasil penelitian menunjukkan bahwa jumlah allel yang didapat adalah 1128. Jumlah allel perlokus berkisar antara 3 (AHT136) sampai 41 (FH2138). Rataan PIC adalah 0,68 dan semua lokus bersifat polimorfik.

Kata kunci : anjing kintamani, mikrosatelit, polymorphic information content (PIC)

Diterima: 11 Agustus 2005, disetujui: 13 Maret 2006

Introduction

The discovery of double helical of DNA brought biology into chemistry and gave new direction to the fields of molecular biology and molecular evolution. Population structure of organism which were originally living based constructed morphological on similarities, have increasingly been assessed from comparisons of DNA. In recent years microsatellite analysis has been widely used to determine population structure within and among population (Koskinen and Bredbacka, 2000; Martines et al., 2000; Stahlberger-Saitbekova et al., 2001).

Microsatellites are repetitive DNA sequence that are randomly distribute throughout vertebrate genomes. Microsatellit repeat sequences for example $(CA)_n$ repeats an well dispersed in the genome, highl polymorphic and have been shown to b powerful tools in genome mapping of dog (Mellersh et al., 2000). They are based on shore 1-5 base repeats and commonly referred to a simple sequence repeats. The repeat motif i most widely distributed in eucaryotic genome is $(dC-dA)_n - (dG-dT)_n$ (Stalling *et al.*, 1991) These microsatellites loci have been shown t be highly polymorphic in the population because of variations in the number of repeat unit (Zajc et al., 1997), with mutation rate often exceeding 10⁻⁴ per generation (Weber and Wong, 1993). Their variability stems from different numbers of basic repeated units. As polymorphic genetics markers, they have been used to improve the accuracy in classification of domestics animals (Nagamine and Higuchi, 2000) and are increasingly becoming the markers of choice for population genetics studies (Bruford and Wayne, 1993). Microsatellites are the best available molecular tools for characterization domestics animal, breed assignment (Canon et al., 2000) and parental assignment (Koskinen and Bredbacka, 1999, Schnabel et al., 2000, Villanueva et al., 2002). They have been used for parentage testing in many species, but the application of microsatellite to parentage testing in dogs is relatively recent (Ichikawa et al., 2001). Microsatellite have been used as one of the most useful tools for genetic identification and to studies of canine pedigrees (Oishi et al., 2005).

Sukawana, a village in a distric of Kintamani, Bangli, province Bali, is an area just 5 km from Kintamani and home to more than 5200 people. Approximately 2100 dogs also live in this area. The Kintamani dogs are highly popular household pets in Bali. The Kintamani dogs have a good appearance and the capacity for circus performances (Puja, 2001). Although Kintamani dogs have a wide range of coat colors, most of the dogs have similar morphological characteristic. The Kintamani dogs are small to medium in size. The wither heights of the female dog is 44.65 cm while the male is 51.21 cm. Kintamani dogs are bold and not aggressive (Puja, 2000). A preliminary studies have been initiated to characterize of Kintamani dogs using craniometry analysis (Puja 2001). This result suggest that the Kintamani dogs are considerably pure. However, the origin of Kintamani dogs remains unknown and this breed is believed to be native to the region of Kintamani, Bali, Indonesia.

The ease of isolation and the utility of microsatellites have made it possible to use them for investigation and apply them to a wide range of different species in which extensive basic genetic analysis has not previously been feasible. As the knowledge

Riota Vol. XI (3) Oktober 2006

about the frequency distribution of microsatellite alleles within canine species currently is sparse, the present study was carried out to characterize of Kintamani dogs using microsatellites markers. Hundreds sixteen canine microsatellite loci, which are known to be polymorphic were chosen from a bank of canine microsatellites.

Materials and Methods

Sample Collection

Buccal swab samples were collected from 425 Kintamani dogs living in their habitat in Kintamani Bali. The samples were taken randomly from individuals from known areas, without consideration of relationship between animals.

DNA Extraction

Genome DNA was extracted from buccal swab cell by heating a single swab for 10 min at 95° C in 400 µl 50mM NaOH and then neutralized with 140 µl 1 M Tris-HCL pH 8.0.

Microsatellite Genotyping

Hundreds sixteen microsatellites were amplified in 12 multiplexes reaction. The multiplexing strategy involves amplifying combinations of markers so that no two markers with the same dye and product size overlap (Cargill et al., 2002). Polymerase chain reaction (PCR) amplification was performed on PCT 100 (MJ Research, Inc, Watertown, Mass, USA) using 30 cycles: denaturation at $95^{\circ}C$ (10 min), annealing at 56°C (30 min) and extension at 72° C (1min). All PCR work was done in polycarbonate 96-well v-bottom microtiters plates. PCR products were run on 6% bis-Acrylamide gel in automated DNA sequencer Prism (ABI 377 DNA Sequencer-PE Biosystem, Foster City, CA, USA). Fluorescent signals from the dye-labeled microsatellites were detected using Genescan 3.1 software (PE Biosystem and Strand Version 2.2.39 program was employed in calculation of the allele number.

I Ketut Puja

Computation

Allele frequency was determined by direct counting and polymorphism information content (PIC) was determined for all markers in each animal of the population.

Result and Discussion

The goal of the present study was to characterize the Kintamani dogs. The microsatellite loci analyzed in the present investigation were selected from a total of 116 sequenced loci. Genetics variabilities within Kintamani dogs population were quantified with microsatellites allel size and polymorphic information content (PIC).

One hundred sixteen primer pairs designed for analyzing canine microsatellites were used for amplification of homologous sequences in Kintamani dogs. A total of 1128 alleles were found in Kintamani dogs. The microsatellite loci revealed that all loci were polymorphic with the number of allele varying from 3 (AHT136) to 41 (FH2138). Frequency distribution and allele size in microsatellite loci in Kintamani dogs are almost the same. The PIC values in Kintamani dogs were 0.68. This is higher than the value of 0.52, calculated by Ostrander and coworkers (1993), and 0.5 by Zajc *et al.*, (1997). PIC might provide a better estimate of the degree of variability, because it depends on the number as well as the frequency of alleles. A considerable reduction in intrabreed population variation was observed in Kintamani dogs population. This result almost certainly reflects high inbreeding coefficient, which varies from breed to breed in domestic dogs. Although the Kintamani dogs were supposedly unrelated, they are probably more inbreed within population.

Population data collected on microsatellites polymorphism in Kintamani dogs can be applied in breed assignment, linkage analysis, studying a segregation of microsatellites and loci associated with quantitative traits or numerous genetics diseases that affect the Kintamani dogs. Screening the canine genome for linkage of markers with various hereditary diseases facilitates identification of affected and carrier individuals, thereby providing researchers an clinicians with an additional diagnostic tool.

This study contributes to the knowledg of genetic structure and molecula characterization of the dog population. It als show how microsatellites can be used t establish the genetic relationship betwee population providing reasonable statistica power for breed assignment, regardless of whether they are closely related or nor allowing their future management to be base on greater knowledge of genetics structurin and relationship between population.

Acknowledgment

This work was supported by Yayasa Yudistira Swarga and by grant from Th University of California, Davis. I gratefull acknowledge Dr.Neil C.Peddersen, Veterinar Genetics Laboratory, School of Veterinar Medicine, UC Davis.

References

- Brufford, M.W. and Wayne, R.K. 1993. Microsatellite and their application to population genetic studies. *Curr Opin Genet Dev.* 3:939-943.
- Canon, J., Checa, M.L., Carleos, C., Vega-Pla, J.L Vallejo, M. and Dunner, S. 2000. The genetics structure of Spanish Celtic hors breeds inferred from microsatellite data *Anim Genet*. 31:39-48.
- Cargill, E.J., Clark, L.A., Steiner, J.M. and Murphy K.E. 2002. Multiplexing of canin microsatellite markers for whole genom screens. *Genomics*. 80:250-253.
- Ichikawa, Y., Takagi, K., Tsumagari, S., Ishiama, K Morita, M., Kanemaki, M., Takeishi, M. an Takahashi, H. 2001. Canine parentag testing based on microsatellit polymorphisms. J Vet Med Sci. 63:1209 1213.
- Koskinen, M.T.and Bredbacka., P. 1999. A convenier and efficient microsatellite-based assay for resolving parentages in dogs.*Anim.Genet* 30 148-149.
- Koskinen, M.T. and Bredbacka, P. 2000. Assessment of the population structure of five Finnish do breeds with microsatellites. *Anim.Gene* 31:310-317.

Genetics Characterization of the Kintamani Dogs

- Martines, A.M., Delgado, J.V., Rodero, A. and Vega-Pla, J.L. 2000. Genetics structure of the Iberian pig breed using microsatellites. *Anim Genet*.31:295-301.
- Mellersh, C.S., Hitte, C., Richman, M., Vignaux, F., Priat, C., Jouquand, S., Werner, P., Andre, C., DeRose, S., Patterson, D.F., Ostrander, E.A. and Galibert, F. 2000. An integrated linkageradiation hybrid map of the canine genome. Mamm. *Genome* 11:120-130.
- Nagamine, Y. and Higuchi, M. 2000. Genetics distances and classification of domestic animals using genetics markers. J.Anim Breed Genet.118:101-109.
- Oishi, N., Maeda, M., Makimura, K., Sawaguchi, T., Hayashiya, M., Kubo, T., Kano, R., Hasegawa, A. and Kasahara, M. 2005. Microsatellite polymorphism in Japanese Mongrel Dogs. J Vet Med Sci.67:1055-1057.
- Ostrander, E.A., Sprague, G.F. and Rine, J. 1993. Identification and characterization of dinucleotide repeat (CA)_n markers for genetic mapping in dog. *Genomic* 16:207-213.
- Puja, I.K. 2000. Maternal behavior in Kintamani bitches during lactation periods. *Media Kedokteran Hewan.* 16: 83-85.
- Puja, I.K. 2001. The estimations of breeds status on Kintamani dogs: Using craniometry analysis. Jurnal Biologi. 5:1-3

- Schnabel, R.D., Ward, T.J. and Derr, J.N. 2000. Validation of 15 microsatellites for parentage testing in North American bison, *Bison bison* and domestic cattle. *Anim Genet*.31:360-366.
- Stahlberger-Saitbekova, Schlapfer, J., Dolf, G. and Gaillard, C. 2001. Genetics relationship in Swiss sheep breeds based on microsatellites analysis. *J.Anim.Breed.Genet*.118-379-387.
- Stalling, R.L., Ford, A.F., Nelson, D., Torney, D.C., Hildebrand, C.E. and Mayris, R.K. 1991. Evolution and distribution of $(GT)_n$ repetitive sequences in mammalian genomes. *Genomic*.10:807-815.
- Villanueva, B., Verspoor and Visscher, P.M. 2002. Parental assignment in fish using microsatellite genetic markers with finite number of parents and offspring. *Anim Genet*.33:33-41
- Weber, J.L. and Wong, C. 1993. Mutation of human short tandem repeats. *Hum Mol Genet*. 2:1123-1128.
- Zajc, I., Cathryn, S., Mellersh and Sampson, J. 1997.Variability of canine microsatellites within and between different dogs breeds. *Mam Genom*.8:182-185.